Understanding the Inflammatory Mechanisms

That Predispose to Emphysema

In Mouse Models

Chuan En Eric Lam

B.BioMed Sc (Hons)

Submitted for the degree of DOCTOR of PHILOSOPHY

Discipline of Immunology and Microbiology
School of Biomedical Sciences and Pharmacy
Faculty of Health
Priority Research Centre for Asthma and Respiratory Disease
The University of Newcastle
Australia

April 2011

Statement of Originality

This thesis contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text. I give consent to this copy of my thesis, when deposited in the University Library, being made available for loan and photocopying subject to the provisions of the Copyright Act 1968.

I hereby certify that the work embodied in this thesis has been done in collaboration with other researchers, or carried out in other institutions. I have included as part of the thesis a statement clearly outlining the extent of collaboration, with whom and under what auspices. This statement can be found on Page III.

Eric Chuan En Lam

April 2011

List of Collaborative Work

This section officially acknowledges the contribution from our collaborators.

Collaborator	Collaboration
A/Prof Philip Hansbro (UoN)	Supply of Non-Typeable <i>Haemophilus influenzae</i> for the investigations carried out in Chapters 3 and 4
Ms. Ama-Tawiah Essilfie (UoN)	Preparation of stock <i>Haemophilus influenzae</i> for infection of mice and bacterial recovery.
Dr. Jodie Simpson (HMRI)	Determining chemerin levels in sputum (comparing between Health and COPD subjects) and analysying results for Figure 4-23 in Chapter 4.

Acknowledgements

I would like to thank my supervisors Dr. Simon Phipps and Prof. Paul Foster for their guidance and support over the last 5 years, as well as the opportunity to undertake this PhD. I would also like to extend my sincere gratitude to Dr. Nicole Hansbro for proofreading the thesis. Special thanks to Adeline for the many great discussions into some of the experimental techniques and great lab mate all round. I would also like to thank all past and present members from the Foster laboratory, who had assisted in one way or another, making this journey a memorable one.

I would like to thank Anne Prins (ANU) and all staff members from the Animal Services Unit (UoN) for the support over the years. I would also like to thank our collaborators A/Prof. Philip Hansbro (UoN) and Ama-Tawiah Essilfie (UoN) for providing the *Haemophilus influenzae* and bacterial recovery work respectively, as well as Dr. Jodie Simpson (HMRI) for supplying the human chemerin data.

I would also like to acknowledge funding support from the University of Newcastle and Cooperative Research Centre for Asthma and Airways for the project.

Last but not least, a huge thank you to my parents, especially my dad for the huge support, encouragement, and funding to see me through this journey. Thank you!

Table of Contents

STATEMENT OF ORIGINALITY	II
LIST OF COLLABORATIVE WORK	III
ACKNOWLEDGEMENTS	IV
TABLE OF CONTENTS	V
ABSTRACT	IX
LIST OF ABBREVIATIONS	XI
CHAPTER 1: INTRODUCTION	1
1.1 Chronic Obstructive Pulmonary Disease	1
1.1.1 Emphysema	5
1.1.2 The Inflammatory response	6
1.1.2.1TNF-lpha	7
1.2 Toll-like receptors	8
1.2.1 Toll-like receptor signalling	11
1.2.2 Synergies between TLR signalling	12
1.2.3 Non pathogen-associated ligands of toll-like receptors	13
1.3 Bacterial infections and COPD	14
1.3.1 Haemophilus influenzae and COPD	15
1.4 Role of apoptosis in emphysema	16
1.4.1 The Regulation of apoptosis	17
1.4.2 Ceramide and Apoptosis	18
1.4.3 Synthesis of ceramide	20
1.4.3.1 Dihydro-Ceramide Synthase (LASS)	21
1.4.3.2 Sphingomyelinase	21
1.4.3.3 Ceramidase	21
1.5 Summary	22
1.6 Hypothesis and Aims	24

CHAPTER 2: LPS AND THE DEVELOPMENT OF EMPHYSEMA "LIKE"			
LESIONS IN THE AIRWAYS OF MICE	26		
2.1 Introduction	26		
2.2 Materials and Methods	29		
2.2.1 Wild type and Genetically modified mice	29		
2.2.2 Mouse models of LPS-induced emphysema	30		
2.2.2.1 An Intratracheal model of LPS-induced emphysema	30		
2.2.2.2 An Aerosol model of LPS-induced emphysema	31		
2.2.2.3 Two week aerosol model of LPS-induced emphysema	31		
2.2.3 Morphometric analysis	33		
2.2.4 Flow Cytometry	36		
2.2.5 Messenger RNA analysis	37		
2.2.5.1 Total RNA extraction	37		
2.2.5.2 Reverse Transcription	38		
2.2.5.3 Primers	39		
2.2.5.4 Analysis of gene expression by quantitative real-time PCR (qPCR)	39		
2.2.6 ELISA	40		
2.2.7 Immunohistochemistry (IHC)	41		
2.2.8 Statistic analysis	43		
2.3 Results	45		
2.3.1 An intratracheal model of LPS-induced emphysema	45		
2.3.2 An aerosol model of LPS-induced emphysema	47		
2.3.3 Temporal analysis of LPS exposure on the development of emphysema	49		
2.3.4 LPS and lung volume	49		
2.3.5 LPS and lung development	49		
2.3.6 Resolution of LPS-induced emphysema	50		
2.3.7 Role of MyD88 in LPS-induced emphysema	54		
2.3.8 Role of MyD88 in LPS-induced inflammation	54		
2.3.9 LPS and CD11c expression on alveolar macrophages	60		
2.3.10 LPS and proinflammatory cytokines in the lung	62		
2.3.11 LPS and proinflammatory chemokines in the lung	63		
2.3.12 LPS and Apoptosis	67		
2.3.13 Age-dependent development of emphysema in TLR-deficient mice	69		
2.4 Discussion	71		
CHAPTER 3: HAEMOPHILUS INFLUENZAE AND THE DEVELOPMEN	T OF		
EMPHYSEMA	85		
3.1 Introduction	85		
3.2 Materials and Methods	87		
3.2.1 Wild type and genetically modified mice	87		

3.2.2 Mouse models of NTHi-induced emphysema	87
3.2.2.1 A single challenge model of NTHi-induced disease	87
3.2.2.2 A double challenge model of NTHi-induced disease	88
3.2.3 Morphometric analysis	89
3.2.4 Flow Cytometry analysis	89
3.2.5 Messenger RNA analysis	89
3.2.5.1 Total RNA extraction	89
3.2.5.2 Reverse Transcription	89
3.2.5.3 Primers	89
3.2.5.4 Analysis of gene expression by quantitative real-time PCR (qPCR)	90
3.2.6 Fluorometric Assay	90
3.2.7 Immunohistochemistry (IHC)	90
3.2.8 Statistic analysis	91
3.3 Results	92
3.3.1 A mouse model of NTHi infection using a single challenge	92
3.3.2 A mouse model of double NTHi inoculation	92
3.3.3 NTHi challenge and lung volume	93
3.3.4 NTHi-induced emphysema and TLR4 signalling	97
3.3.5 NTHi challenge and the inflammatory response	99
3.3.6 NTHi challenge and the expression of proinflammatory cytokines	99
3.3.7 NTHi challenge and the expression of proinflammatory chemokines	102
3.3.8 NTHi challenge and activation of the apoptotic pathway	102
3.3.9 NTHi challenge and the synthesis of ceramide	103
3.4 Discussion	107
CHAPTER 4: NEW APPROACHES TO INHIBIT THE DEVELOPMEN	T OF LPS
OR HAEMOPHILUS INFLUENZAE-INDUCED EMPHYSEMA	115
OK HAEMOFHILUS INFLUENZAE-INDUCED EMFILISEMA	113
4.1 Introduction	115
4.1.1 Chemerin	117
4.1.2 Chemerin receptors	118
4.1.2.1 Orphan chemerin receptor - ChemR23	118
4.1.2.2 Orphan chemerin receptor - CCRL2	119
4.1.3 Chemerin-derived 15-amino acid (C15)	119
4.1.4 Inhibiting inflammation	120
4.2 Materials and Methods	122
4.2.1 WT mice	122
4.2.2 Peptide synthesis	122
4.2.3 Mouse models and treatment	123
4.2.3.1 Determining chemerin levels	123
4.2.3.2 Optimizing dose of old SP and C15	123
4.2.3.3 Determination of the optimal route of administration of SP and C15	125
4.2.3.4 Prophylactic administration of SP and C15	125

R	EFERENCES	200
C	HAPTER 5: GENERAL DISCUSSION AND CONCLUSIONS	181
4.	4 Discussion	168
	4.3.20 Chemerin levels in human patients	166
	4.3.19 The role of alveolar macrophages in a mouse model of LPS-induced emphysema	162
	4.3.18 Effect of exposure to a combination of LPS and NTHi	160
	the 2 week LPS aerosol model	158
	4.3.17 Effect of therapeutic administration of C15 on ceramide production related enzym	es in
	4.3.16 Effects of therapeutic administration of C15 on LPS and NTHi-induced emphysema	156
	4.3.15 Effect of C15 on bacterial clearance	153
	4.3.14 Effect of C15 on LPS- or NTHi-induced caspase-3 activity in the lung	153
	4.3.13 Effect of C15 on LPS- or NTHi-induced ceramide levels in alveolar macrophages	151
	4.3.12 Effect of C15 on NTHi-induced enzymes associated with ceramide synthesis	148
	4.3.11 Effect of C15 on LPS-induced enzymes associated with ceramide synthesis	148
	4.3.10 Effect of C15 on NTHi-induced proinflammatory chemokines	145
	4.3.9 Effect of C15 on LPS-induced proinflammatory chemokines	145
	4.3.8 Effects of C15 on Chemerin expression and associated receptors	141
	4.3.7 Effect of C15 and SP on LPS- and NTHi-induced inflammation	140
	4.3.6 Effect of C15 and SP on lung volumes	140
	4.3.5 Effects of C15 on LPS and NTHi-induced emphysema	136
	4.3.4 Effect of C15 on LPS-induced emphysema in older mice	136
	4.3.3 Routes of administration of peptides (SP or C15) affect its efficacy	134
	4.3.2 New scrambled peptide (SP) exerts no effects on LPS or NTHi treatment	132
4.	4.3.1 Chemerin levels in LPS- or NTHi- treated mice	130
1	.3 Results	130
	4.2.10 Statistic analysis	129
	4.2.9.2 Human Chemerin	129
	4.2.9.1 Murine Chemerin	129
	4.2.9 Protein detection via ELISA	128
	4.2.8 Immunohistochemistry	128
	4.2.7 Fluorometric Assay	128
	4.2.6.4 Analysis of gene expression by quantitative real-time PCR (qPCR)	128
	4.2.6.3 Primers	128
	4.2.6.2 Reverse Transcription	128
	4.2.6.1 Total RNA extraction	128
	4.2.6 Messenger RNA analysis	128
	4.2.5 Flow Cytometry analysis	128
	4.2.4 Morphometric analysis	128
	4.2.3.5 Therapeutic administration of SP and C15	127

Abstract

Chronic obstructive pulmonary disease (COPD) is a growing global health problem, and this disorder is projected to rank fifth by 2020 as a worldwide burden of disease (Murray and Lopez., 1996). Remarkably, little is known about the pathogenesis of COPD and current pharmacologic agents fail to halt disease progression. Emphysema is a major inflammatory disorder that falls under the clinical description of COPD. Emphysema can be induced by smoking but can also occur in non-smokers. Emerging data suggests that the loss of alveolar tissue which characterises emphysema may result from increased cell death (apoptosis) of alveolar epithelial cells mediated by the sphingolipid mediator ceramide (Petrache et al., 2005). The cause of COPD exacerbations are commonly bacterial or viral respiratory infections. Under certain conditions, immunity from infection is mediated through the initiation of apoptotic pathways by infected cells to prevent the pathogen from replicating within the host. Toll-like receptors (TLRs) recognise molecular patterns expressed by pathogens such as bacteria and viruses to initiate innate immune responses. Notably, significant amounts of the bacterial wall component lipopolysaccharide (LPS) are found in cigarette smoke. LPS is a TLR4 ligand that increases the level of the apoptotic mediator ceramide and production of proinflammatory cytokines (such as tumour necrosis factor $(TNF)-\alpha$, interleukin (IL)-1 β , and IL-6) implicated in the pathogenesis of emphysema. We hypothesise that chronic inhalation of LPS leads to the dysregulation of TLR4 signalling pathways that increases susceptibility to

respiratory infection, and uncontrolled inflammation that promotes alveolar cell apoptosis and emphysematous-like lesions. We developed mouse models of LPSand bacterial-induced emphysema to determine if attenuating inflammation can prevent the development of emphysema.

Our results demonstrate that exposure to LPS or infection with Nontypeable Haemophilus influenzae (NTHi) (often found in patients with emphysema) can induce hallmark features of emphysema, such as alveolar enlargement (determined by mean linear intercept and percentage alveolar airspace measurements) and inflammation dominated by neutrophils and macrophages. We demonstrated that alveolar enlargement was due to the loss of alveolar parenchyma (from apoptosis), is dependent on TLR4 and myeloid differentiation factor-88 (MyD88), increased proinflammatory cytokines, chemokines, and inflammatory cells (neutrophils and macrophages) in the lung. Prophylactic administration of synthesised chemerin-derived peptide (C15) attenuated LPS- or NTHi-induced inflammation, which resulted in inhibition of the development of emphysematous-like lesions. Notably, specific depletion of alveolar macrophages protects mice from LPS- or NTHi-induced emphysema.

Collectively, we demonstrate that blocking inflammation during the development of emphysema is critical for preventing or attenuating the progression of the disease.

List of Abbreviations

AA Amino acid

ALF Australian Lung Foundation

APAAP Mouse Alkaline-Phosphatase anti-Alkaline-Phosphatase

Apaf-1 Apoptotic protease activating factor-1

APCs Antigen presenting cells
ASmase Acid Sphingomyelinase
CAD Caspase-activated DNAse

CCRL2 Chemokine (CC motif) receptor-like 2

cDNA complementary DNACKRX Chemokine Receptor XCMKLR1 Chemokine-like receptor 1

COPD Chronic obstructive pulmonary disease

2CA 2-ChloroadenosineECM Extracellular matrixE.coli Escherichia coli

EtOH Ethanol

FCS Fetal Calf Serum

FEV₁ Forced expiratory volume in one second

FVC Forced vital capacity

GOLD Global initiative for Chronic Obstructive Lung Disease

GPCR Protein-coupled receptors

H.influenzae Haemophilus influenzae

HKR Human chemokine receptor

HMGB-1 High-mobility group protein B1

HMRI Hunter Medical and Research Institute

IRFs Interferon regulatory factorsIVC Individually ventilated cagesLASS Dihydroceramid Synthases

L-CCR LPS-inducible C-C chemokine receptor related gene

LOS Lipooligosaccharides
LPS Lipopolysaccharide
MAL MyD88 adaptor-like

MiP-1α Macrophage inflammatory Protein -1 -alpha

MLI Mean linear intercepts
MMP-9 Matrix metallopeptidase 9
MMP-12 Macrophage Elastase
mRNA messenger RNA
MyD88-^{-/-} MyD88-deficient

MyD88 Myeloid-Differentiating factor 88

NADPH reduced Nicotinamide adenine dinucleotide phosphate

NCdase Neutral Ceramidase
NGS Normal Goat Serum

NHLBI National Heart, Lung, and Blood Institute

NLRs NOD-like receptors Nox-3 NADPH oxidase 3

NTHi Non-typeable Haemophilus influenzae
PAMPs Pathogen-associated molecular patterns

PBS Phosphate Buffered Saline

PFA Paraformaldehyde

% alveolar airspace (%AA) percentage alveolar airspace

qPCR quantitative real-time PCR

RARRES2 Retinoic acid receptor responder 2

RSV Respiratory Syncytial Virus

SARM Sterile α and armadillo motif-containing protein

SMases Sphingomyelinases

SMS Sphingomyelinase Synthase

SP Scrambled Peptide

S. pneumoniae Streptococus pneumoniae

S1-P Sphingosine-1-phosphate

Tg Transgenic

TIG2 tazzarotene induced gene 2

TIR Toll-IL-1 receptor
TLR Toll-like receptor
TLR4-/TLR4 deficient

TNF-α Tumour necrosis factor-alpha

TRAIL TNF-related apoptosis inducing ligand

TRAM TRIF-related adaptor molecule

TRIF TIR domain-containing adaptor inducing interferon-β
TSANZ The Thoracic Society of Australia and New Zealand

VEGF Vascular Endothelial Growth Factor

WHO World Health Organisation

WT Wild type